

quantitative conversion to 5-hydroxypipelicolic acid. Strong but not conclusive evidence has been adduced that both the *cis* and *trans*-5-hydroxypipelicolic acids are present in *G. triacanthos*. Determination of the iminoacids was established by co-chromatography.

EXPERIMENTAL

Seeds of *G. dioicus* (L) Koch. were collected in October 1972 in Oklahoma (USA). Voucher specimens are deposited in the Department of Organic and Biological Chemistry, Faculté des Sciences Agronomiques, Gembloux.

Isolation of *cis* and *trans* 5-hydroxypipelicolic acid. (For extraction, purification and separation of acidic, neutral and basic amino acids, see ref. [2]). Neutral extract from 1.7 kg of seeds was dissolved in a min. vol. of 1.5 N HCl and passed through a column (150 × 5 cm) of Lewatit S 1080 in the H⁺ form. The amino acids were eluted with 1.5 N HCl (12 ml fractions). 5-Hydroxypipelicolic acid is eluted in fractions 80–140 (9 g) and pipelicolic acid in fractions 160–195. *Cis* and *trans*-5-hydroxypipelicolic were then separated on Dowex 50 W × 8 (50 × 4 cm) in the Na⁺ form, equilibrated with a buffer sol. of pH 3.2 [2]. *Cis*-5-hydroxypipelicolic acid is eluted in fractions 235–250 and *cis* + *trans*-5-hydroxypipelicolic acid in fractions 251–335 (10 ml fractions). From the latter fractions, *cis* and *trans*-5-hydroxypipelicolic acids were separated using preparative PC (*tert* amyl alcohol-2,4-lutidine-H₂O; 89:89:57). Elution was performed during 5 days. $R_{cis/trans} = 0.84$ (Piez *et al.*: 0.83) [21]. *Cis*-5-hydroxypipelicolic acid recrystallized (170 mg): C, 49.65; H, 7.64; N, 9.65 calculated for C₆H₁₁NO₃: C, 49.47; H, 7.61; N, 9.62%. $[\alpha]_{589}^{22} = -29.5^\circ$ (c, 2; H₂O), $[\alpha]_{589}^{22} = -17.3^\circ$ (c, 1; HCl 6 N). IR(KBr) $\nu_{max}^{cm^{-1}}$: 3400 (large), 3000 (large), 1625 (s), 1440(w), 1400(s), 1340(w), 1315(w), 1210(w), 1165(w), 1130(w), 1090(m), 1040(m), 980(m), 935(w), 850(w), 770(m).

General instrumentation. PC was performed in *n*-BuOH-HOAc-H₂O (12:3:5) and pH 4.2. HVE was carried out in buffer at pH 1.9; 60 V/cm; 90 min. The mobilities of the hydroxypipelicolic acids in cm were *cis* form = 13.6; *trans* form = 12.0 PMR spectra were measured at 100 MHz and chemical shift are in ppm downfield from 2,2,3,3-tetra-deuterio-3-(tris-TMS)propionate in D₂O.

From a personal communication, we have just heard that Hatanaka has isolated *cis*-5-hydroxypipelicolic acid from *Lathyrus japonicus* (seeds) and *cis* and *trans*-5-hydroxypipelicolic acid from *Morus multicaulis* (leaves).

Acknowledgements—The authors express their gratitude to Professor E. A. Bell, King's College, London for providing seeds. They wish to thank Professor J. Casimir for valuable discussions during the present work. They are indebted to the Facultés Universitaires de Namur (Professor Krief) for providing facilities for PMR spectrometry.

REFERENCES

1. Dardenne, G. (1970) *Phytochemistry* **9**, 924.
2. Dardenne, G., Casimir, J., Bell, E. A. and Nulu, J. R. (1972) *Phytochemistry* **11**, 787.
3. Dardenne, G., Bell, E. A., Nulu, J. R. and Cone, C. (1972) *Phytochemistry* **11**, 791.
4. Evrard, G., Durand, F. and Dardenne, G. (1974) *Cristal. Struct. Commun.* **3**, 65.
5. Dardenne, G., Casimir, J. and Sørensen, H. (1974) *Phytochemistry* **13**, 2195.
6. Grobbelaar, N., Pollard, J. K. and Steward, F. C. (1955) *Nature* **175**, 703.
7. Witkop, B. and Foltz, C. M. (1957) *J. Am. Chem. Soc.* **79**, 192.
8. Clark-Lewis, J. W. and Mortimer, P. I. (1961) *J. Chem. Soc.* 189.
9. Schenk, W. and Schütte, H. R. (1963) *Flora* **153**, 426.
10. Hatanaka, S. (1969) *Phytochemistry* **8**, 1305.
11. King, F. E., King, T. J. and Warwick, A. J. (1950) *J. Chem. Soc.* 3590.
12. Marlier, M., Dardenne, J. and Casimir, J. (1972) *Phytochemistry* **11**, 2597.
13. Evrard, G., Durand, F. and Marlier, M. (1972) *Cristal. Struct. Commun.* **1**, 215.
14. Marlier, M., Dardenne, G. and Casimir, J. (1976) *Phytochemistry* **15**, 183.
15. Shewry, P. R. and Fowden, L. (1976) *Phytochemistry* **15**, 1981.
16. Kristensen, I., Larsen, P. O. and Sørensen, H. (1974) *Phytochemistry* **13**, 2803.
17. Dardenne, G., Casimir, J. and Sørensen, H. (1974) *Phytochemistry* **13**, 1515.
18. Hatanaka, S. (1972) *College of Gen. Educ. Univ. of Tokio* **22**, 2, 117.
19. Despontin, J. (1976) Thesis, Faculté des Sciences Agronomiques, Gembloux.
20. Thompson, J. F. and Morris, C. J. (1968) *Arch. Biochem. Biophys.* **125**, 362.

Phytochemistry, 1977, Vol. 16, pp. 388–390, Pergamon Press, Printed in England

SOLUBLE SUGARS OF MAIZE SEEDLINGS*

J. A. KALAITZAKIS, N. S. MARGARIS† and C. A. THANOS

Institute of General Botany, University of Athens, Athens 621, Greece

(Received 9 August 1976)

Key Word Index—*Zea mays*; Gramineae; maize seedlings; soluble sugars.

Abstract—The soluble sugars were determined in different parts of maize seedlings (seeds, roots and shoots), 0, 2, 4 and 6 days after sowing.

INTRODUCTION

Although germination, as a series of biochemical processes, has been extensively studied, information on

such 'simple' biochemical events as the hydrolysis of seed polymers, is still lacking [1]. The present work approaches the chemical differences during early stages